



# MULTICENTER STUDY ON PROGNOSTIC AND PREDICTIVE IMPACT OF TP53 VARIANTS BELOW 10% VAF: Update

*49th General Assembly of ERIC Members at EHA*

*25th EHA Annual Congress*

*June 11, 2020*

## BACKGROUND

Should we decrease 10% (5%) cut-off for reporting TP53 mutations?  
If yes, how much?

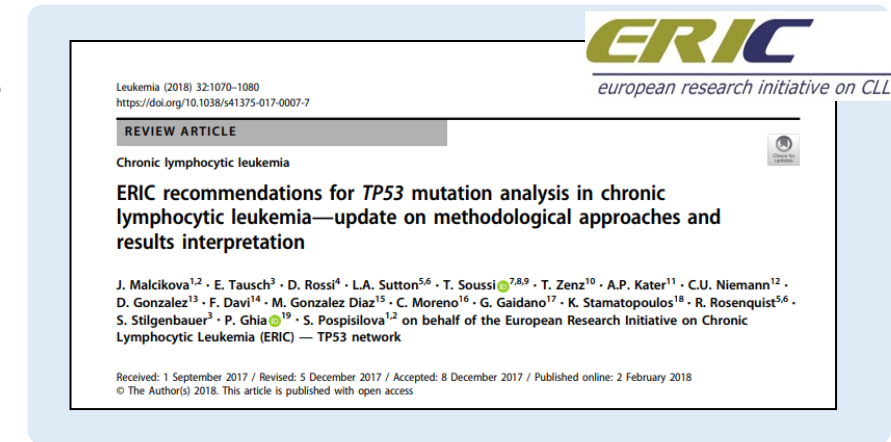
## PROJECT AIMS

**TECHNICAL ASPECT – WHAT IS A SAFE CUT-OFF FOR RELIABLE DETECTION IN ROUTINE DIAGNOSTICS?**

- To compare NGS results among laboratories performing NGS detection of TP53 mutations in CLL with detection limit of 1% VAF

**CLINICOBIOLOGICAL ASPECT – WHAT IS THE LOWEST SIZE OF MUTATED CLONE THAT BRINGS HARM TO THE PATIENT IF NOT PROPERLY TREATED?**

- To confirm prognostic impact of low-VAF TP53 variants in patients entering first-line treatment, both for unmut-IGHV and mut-IGHV



## AIMS

### TECHNICAL ASPECT – WHAT IS A SAFE CUT-OFF FOR RELIABLE DETECTION IN ROUTINE DIAGNOSTICS?

- To compare NGS results among laboratories performing NGS detection of *TP53* mutations in CLL with detection limit of 1% VAF

### CLINICOBIOLOGICAL ASPECT – WHAT IS THE LOWEST SIZE OF MUTATED CLONE THAT BRINGS HARM TO THE PATIENT IF NOT PROPERLY TREATED?

- To confirm prognostic impact of low-VAF *TP53* variants in patients entering first-line treatment, both for unmut-IGHV and mut-IGHV

## HOW TO

### PHASE 1:

Inter-laboratory comparison of NGS results obtained from the set of reference samples =  
**METHODICAL HARMONIZATION**

### PHASE 2:

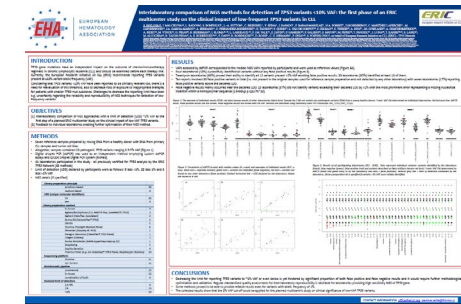
**DATA COLLECTION - results of TP53 analysis using NGS with detection limit ~1% VAF**

Required data: Results of TP53 analysis using NGS methodology with detection limit ~1% VAF.

Corresponding clinical and laboratory data (disease course and routine prognostic markers)

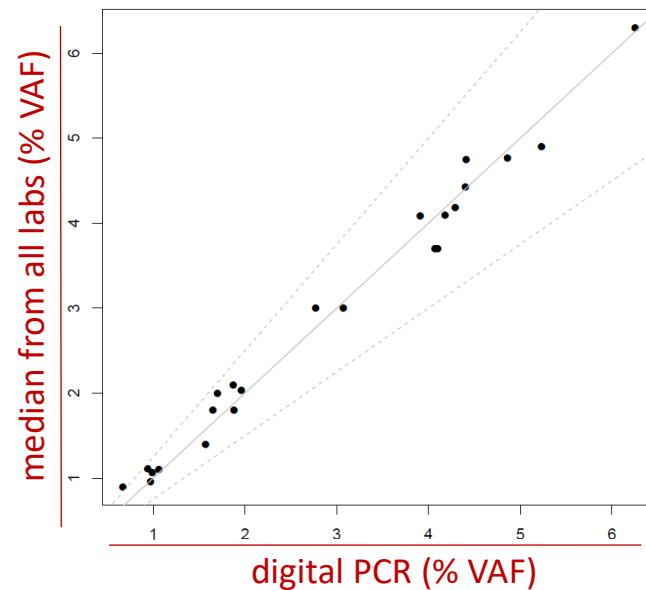
# RESULTS OF PHASE 1:

## Inter-laboratory comparison of NGS results obtained from the set of reference samples



sample	variant (c)	variant (p)	ddPCR [%]
mTP53_1	c.166G>T	p.(E56*)	3.9
	c.524G>A	p.(R175H)	4.3
	c.580C>T	p.(L194F)	4.1
	c.743G>A	p.(R248Q)	2.8
	c.853G>A	p.(E285K)	4.2
mTP53_2	none	none	-
mTP53_3	c.166G>T	p.(E56*)	0.9
	c.524G>A	p.(R175H)	1.1
	c.580C>T	p.(L194F)	1.0
	c.743G>A	P.(R248Q)	0.7
	c.853G>A	p.(E285K)	1.0
mTP53_4	c.173del	p.(P58Qfs*65)	6.3
	c.949dup	p.(Q317fs*20)	5.2
mTP53_5	c.559+1G>A	p.?	1.7
mTP53_6	c.173del	p.(P58fs*65)	1.9
	c.949dup	p.(Q317fs*20)	1.6
mTP53_7	c.337T>G	p.(F113V)	2.0
	c.569C>T	p.(P190L)	4.4
	c.626_627del	p.(R209fs*6)	4.1
	c.672+1G>T	p.?	4.4
	c.685_689del	p.(C229fs*9)	3.1
	c.741_742delinsTT	p.(R248W)	4.9
	c.817C>T	p.(R273C)	1.9
	c.949C>T	p.(Q317*)	1.7

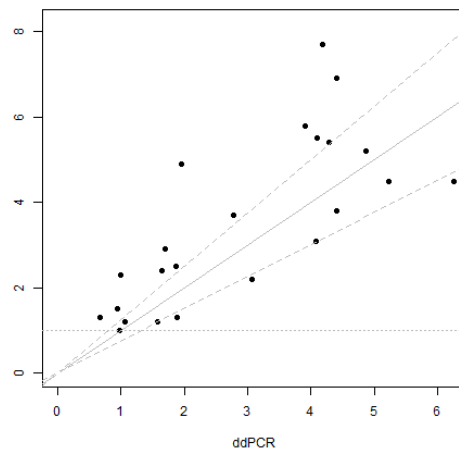
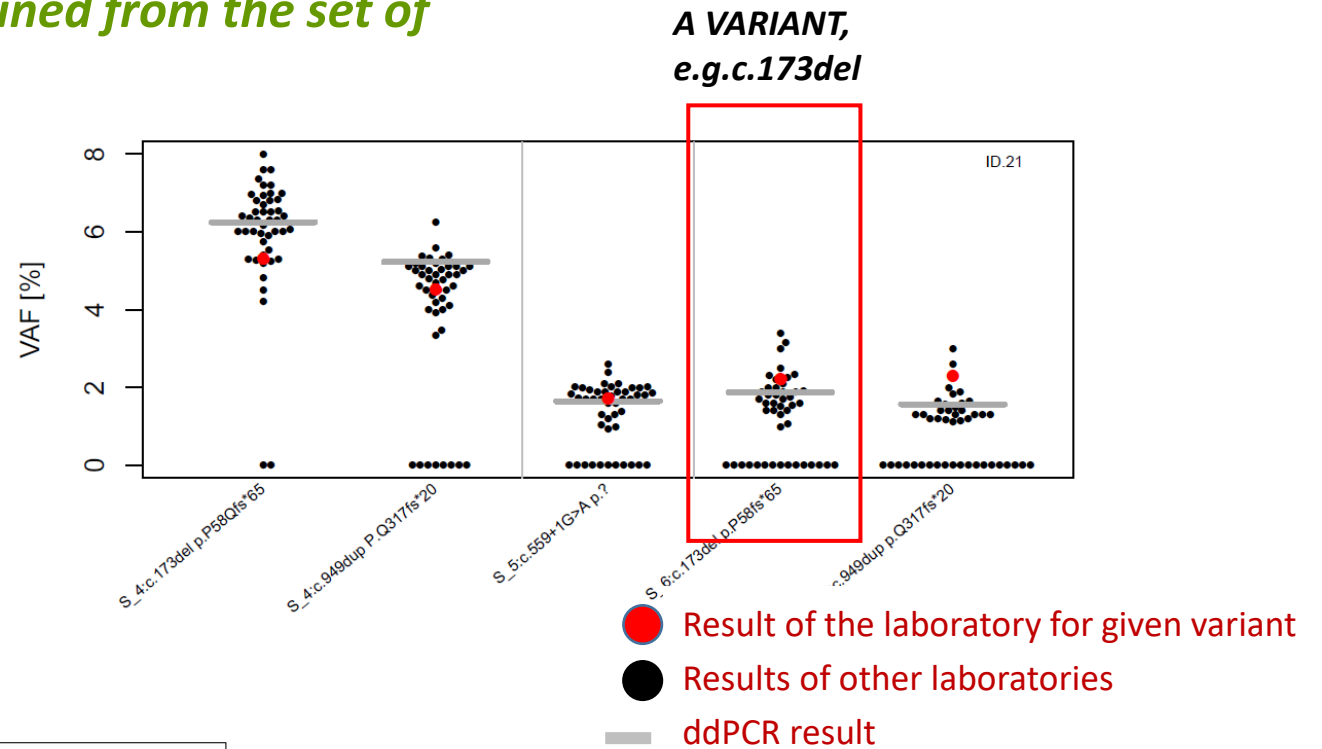
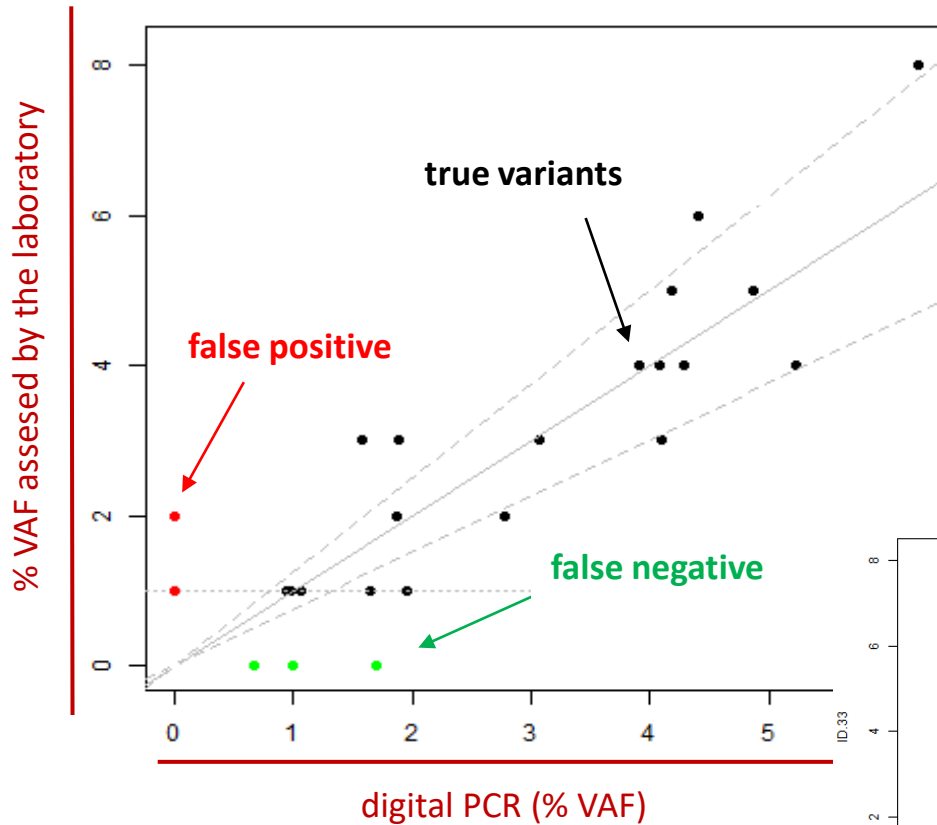
- 7 samples with 23 variants 0.7 – 6.3 % VAF
- 43 participants from 18 countries registered, 41 sent results
- Wide range of commercial and *in-house* NGS approaches
- Limit of detection 1% prevailed
- VAF assessed independently by ddPCR: good correlation with median values



2018 November 1: Invitation  
 2019 May 10: Samples sent  
 2019 May 23: 1<sup>st</sup> results  
 2019 November 20: Last results  
 2020 March 1: EHA abstract  
 2020 June 10: Personalized reports

# PHASE 1: PERSONALISED REPORTS

Inter-laboratory comparison of NGS results obtained from the set of reference samples

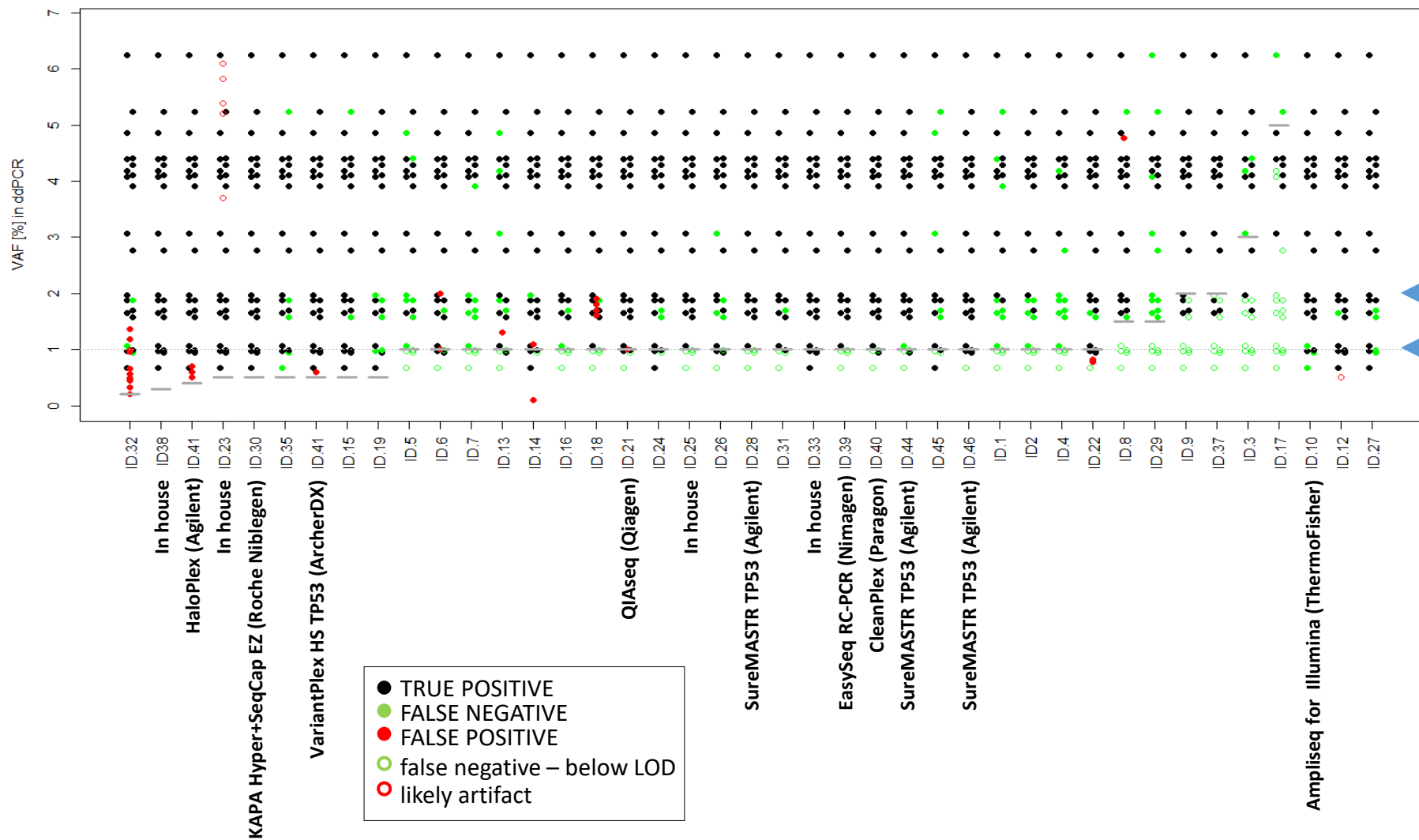




# PHASE 1: OVERALL RESULTS

## Inter-laboratory comparison of NGS results obtained from the set of reference samples

41 LABORATORIES



23 VARIANTS (0.5-7% VAF)

Cut-off	Total*	Not found*	False pos*
2% VAF	396	5.6%	1.0%
1% VAF	627	13.2%	3.3%

\*only laboratories declaring LOD ≤1% VAF (N=33) were considered

## PHASE 2: WHAT`S COMING NEXT?

DATA COLLECTION - results of TP53 analysis using NGS with detection limit <sup>2%</sup>~~~1%~~ VAF

1. Do you wish to participate in the data collection part of the ERIC study? This would require to provide data from a consecutive set of patients entering first-line therapy before 2016 analyzed for TP53 mutations by a reliable NGS methodology with a threshold  $\leq 2\%$  VAF.
2. If yes, which deadline is feasible for you to provide the data?
3. If yes, how many patient samples do you plan to include, approximately?
4. In the case that the harmonization phase showed some shortcomings of your NGS approach, are you able to improve it?

NGS results: variants, VAF

Disease course: dates of diagnosis, therapy, follow-up

Laboratory data: IGHV, FISH

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